An integrated approach to the origin and diversification of protostomes

Goal 1:

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http://www.mcz.harvard.edu/Departments/InvertZoo/tol/



- Generate a collection of tissues for 265 species belonging to all protostome phyla. Having such a collection available for research will ensure gathering of different sources of data for the same organisms across laboratories worldwide,
- Generate sequence data (about 10 markers that have shown to resolve metazoan deep divergences) for 50% of the metazoan lineages proposed in the previous section. Generate new cDNA libraries for 8 protostomes.
- Generate sequence data for about 100 molecular markers by EST technology for the ca. 15 taxa for which the cDNA libraries are generated/available
- Generate new embyrological cell lineage studies for a selected collection of protostome phyla for which cDNA libraries have been generated.
- Generate gene expression data for several genes implied in segmentation, mesoderm, and body plan organization. Compile a comprehensive data set on protostomes morphology/anatomy coded for exemplar taxa, including key fossil species (see accessory Table).



- and Sweden Tissues of more than 200 species belonging to Accela, Xenoturbellida, Myzostomida, Nemertea, Mallusca, Annelida, Sipuncula, Entoprocta, Brachiopoda, Phoronida, Platyhelminthes, Grathostomulida, Gastrotricha, Rottlera, Cycliophora, Micrognathozoa, Priapula, Kinorhyncha, Tardigrada, Loricifera, Onychophora, Arthropoda have been collected and preserved in RNAlater, 96% EtOH, formalin, and alutaralde
- Online database of species, modes of preservation, and status of work is under development. A preliminary version can be viewed at: http://collections.oeb.harvard.edu/Invertebrate/ato//species.cfm

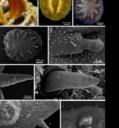


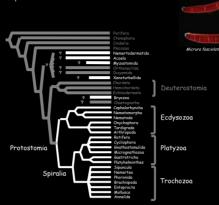
Xenaturbella backi dredaed in Tiärno. Sweder

- n and Florida, Clockwise: Fred Pleijel, Greg Rouse eberg: F. Pleijel and Akiko Okusu with the skipper trineberg: Martin Sorenten collection cost
- Study case: An unknown protostome species is discovered and studied; The "guasi-model" approach
- Description and morpho-anatomical characterization of animal (light microscopy, SEM, TEM)
- Spawning for study of larvae and developmental stages Generation of cDNA library
- Sequencing of ca. 1,400 ESTs Study of expression patterns

We will apply this integral approach to a number of key species to understand protostome evolution, including Xenoturbella accels, and others







Working hypothesis of metazoan relationships highlighting the protostomes and other putative protostomes covered in this study

Goal 2:

- Markers that amplify via PCR or rtPCR 185 rRNA (ca. 1.8 Kb)
 - 285 rRNA (ca. 3 Kb)

 - Myosin heavy chain II (ca. 800 bp) RNA Polymerase TT
 - Elongation factor 1a
 - Elongation factor 2
 - Histone H3 (cg. 350 bp)
 - Cytochrome c oxidase subunit I (ca. 1,200 bp)

We are sequencing other more traditional markers via PCR or rtPCR amplification.

Goals 3 and 4:

- cDNA libraries have been built for 5 protostomes: Myzostoma seymourcollegensis n. sp. (Myzostomida) Cerebratulus lacteus (Nemertea) Themiste lageniformis (Sipuncula) Xenoturbella bocki (Xenoturbellida Phoronis vancouverensis (Phoronida)
- ESTs have been sequenced for 5 animals: Mnemiopsis leidyi (Ctenophora), 1,000 clones Capitella sp. I (Annelida), 1,000 clones caprelia sp. 1 (Anneliad), 1,000 clones Myzostoma seymourcollegensis n. sp. (Myzostomida), 1,400 clones *Cerebratulus lacteus* (Nemertea), 900 clones *Themiste lageniformiis* (Sipuncula), 100 clones (in progress) Xenoturbella backi (Xenoturbellida), 3,000 clones (in progress)

We have obtained homologues of several markers that could be useful for phylogenetics such as tropomyosin, elonaation factors, a-ubulin, Dynein, Notch, myosines, etc.

Brief protocol: Specimens are collected in the field, preserved in RNAlater, transferred to Kewalo for generation of cDNA, plates are sent to the sequencing facilities of the AMNH or Harvard.





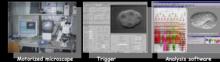




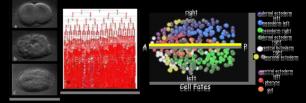
The AToL-protostome team met at Harvard, January 2004. A second meeting is scheduled at Kewalo, January 2005.

Goal 5:

Cell lineages for selected species of metazoans. An example from the gastrotrich Lepidodermella squamata. A new technique using a 4D microscope has been developed to study cell lineages in small invertebrates. With this technique we are able to expand the number of embryos that can be studied for cell lineages without the hassle of cell injections



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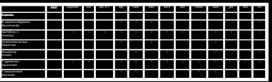




Goal 6:

Generate expression patterns of developmental regulatory genes potentially involved in segmentation, body plan formation and mesoderm formation

The following genes have been rescued so far:



In situ hybridization of Capitella sp. I, ventral view:



The state of the state

hows a very complex pat n some of the mesoderm

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-gsx -xlox

parahox





